

Taconic Assay Transfer Sample Submission Form

I. Contact Information

Institution:

Principle Investigator:

Phone:

Email:

Genotyping Contact:

Phone:

Email:

II. Billing Information

Institution Name:

Address:

City: State: Zip Code:

Accounts Payable Contact Information – Name:

Phone Number: Email:

III. Animal Model Information

Full model name:

Are there multiple manipulations (genes) for this model?

If yes, please provide information below on each gene of interest.

If yes, can the assays for each gene be multiplexed?

Type of Alleles to Identify: (check all that apply)

- | | | |
|---|---------------------------------------|--|
| <input type="checkbox"/> Transgenic | <input type="checkbox"/> Knockin | <input type="checkbox"/> Knockout/Null |
| <input type="checkbox"/> Conditional | <input type="checkbox"/> Constitutive | <input type="checkbox"/> Humanization |
| <input type="checkbox"/> RNAi Knockdown | | |

Please select the mutation method for the animal model in question and answer the corresponding questions:

A. Targeted Mutation Knockout or Knockin

Please specify deletion(s) [exons and introns]:

Where is the insertion site?

Where are the primers located?

Was Neo Hygro or Puro or other just inserted?

Was any sequence replaced with Neo, Hygro, Puro or other?

B. Transgene

What vector(s) were used?

Where is the insertion site?

Was cDNA or genomic DNA used?

What is the copy number?

C. Point Mutation

What is the nucleotide that changes?

Provide the surrounding sequence (~ 200bp on each side):

Is it a spontaneous or constructed point mutation?

D. Conditional Mutation

Was a Cre Used?

Name of Cre line:

Original Cre Line Promotor:

Reference for Cre Model:

Tissues Where Cre is expressed:

Can Cre expression resulting germline event?

Are both the floxed and the null alleles present?

Is the presence of all 3 alleles possible in the tail sample?

Is the null genotype detectable in tail tissue?

Are there plans to use a Fip assay as well?

IV. Assay Information

Primers: (please provide names and sequences for all primers used in PCR reactions)

Primer Name:

Sequence: 5'

Primer Name:

Sequence: 5'

Primer Name:

Sequence: 5'

Primer Name:

Sequence: 5'

Probes: (if applicable)

Sequence and fluorescent moiety, label and location

Probe Name:

Sequence: 5'

Expected PCR Product Sizes: (base pairs)

Homozygote:

Wild type:

Transgene:

Floxed:

Null:

If available please include construct diagrams, vector diagrams, sequence of the insert site and any journal article that pertains to the construction of the modified animal.

Please note:

It is the policy of Taconic to perform an initial study to validate the efficacy and utility of an assay prior to accepting production samples. Due to Taconic's approach to high-throughput PCR genotyping, our scientific professionals will develop the genotyping assay using pre-selected reagents and the client-submitted protocol as a guideline **only**. If the transfer is unsuccessful (i.e. results not reproducible), Taconic will contact the client as to how to proceed before additional work is performed. Additional charges may result if the assay cannot be readily transferred with reproducible results.

Please submit this completed form to molecular.analysis@taconic.com